

Tumor Vasculature as a Therapeutic Target in Non-small Cell Lung Cancer

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Introduction: We aim to describe the molecular mechanisms relevant to angiogenesis inhibition and to critically evaluate the current evidence for the use of angiogenic inhibitors (AIs) in the treatment of non-small cell lung cancer (NSCLC).

Methods: The literature on the basic concepts of tumor angiogenesis is reviewed. Published articles and major lung cancer conference abstracts were screened for reports on the use of AI in NSCLC patients and the National Institutes of Health clinical trials database was searched for relevant ongoing studies.

Results: We delineate in this review the molecular and cellular aspects of angiogenesis and vasculogenesis and outline the relevance of these to lung cancer. Clinical studies of AIs in NSCLC reported to date as well ongoing studies are summarized. Major issues discussed include the choice of the right molecular target; characteristics of various tyrosine kinase inhibitors; potential drawbacks and concerns regarding the application of AIs in clinical practice, and major unanswered questions and future directions.

Conclusions: AIs have antitumor activity in NSCLC and have become part of the standard of care for patients with advanced nonsquamous cell carcinoma. Unfortunately, the gains have been modest and robust predictive biomarkers are urgently needed. Clinical trials to date have validated the tumor vasculature as a legitimate target, and as our understanding of the biology of tumor angiogenesis increases, exciting new therapeutic approaches are being explored.

Key Words: Angiogenic inhibitors, Non-small cell lung cancer, Vascular endothelial growth factor, Tyrosine kinase inhibitors, Predictive biomarkers.

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Lung Cancer Angiogenesis

As a malignant growth exceeds the size of a few 100 μm , nutrient diffusion becomes a growth-limiting factor. Hypoxia and cancer-specific genetic abnormalities, mainly by up-regulation of the hypoxia inducible factors, drive the

secretion of proangiogenic factors (e.g., vascular endothelial growth factor [VEGF], basic fibroblast growth factor [FGF]) and the suppression of antiangiogenic factors (e.g., thrombospondin, endostatin), making the tumor microenvironment proangiogenic. New blood vessels are formed, and existing blood vessels are modified to provide blood supply to the tumor. The sprouting of blood vessels from existing blood vessels is called angiogenesis, whereas production of de novo blood vessels is termed vasculogenesis. Both processes are controlled by the counteracting effects of proangiogenic and antiangiogenic factors. The tipping of the balance toward a proangiogenic state, “the angiogenic switch,” is essential for cancer progression. Unlike in physiological angiogenesis, reperfusion and reoxygenation do not turn off cancer angiogenesis, which is driven by tumor-secreted factors. Tumoral angiogenesis was suggested as a therapeutic target 40 years ago.¹ However, only recently have angiogenesis inhibitors been added to the available therapeutic armamentarium against colon, breast, kidney, brain, and other cancers. Figure 1 depicts some of the major cellular and molecular factors in cancer angiogenesis.

Histological evidence of enhanced angiogenesis in lung tumors has been associated with a poor prognosis^{2,3} and levels of various molecular mediators of the angiogenic switch correlate with poor clinical outcome.⁴ The clinical importance of angiogenesis inhibition in the treatment of non-small cell lung cancer (NSCLC) has been demonstrated.⁵ However, it should be noted that a subgroup of NSCLC with nonangiogenic pattern has also been described.^{6–8} These tumors seem to co-opt existing blood and lymphatic vessels rather than induce angiogenesis, and importantly have a worse prognosis than their angiogenic counterparts. Hence, it is unlikely that antiangiogenic treatments would be effective for all lung cancer patients.

Major Molecular Mediators of Angiogenesis

Angiogenesis involves multiple cellular events and many interactions among a variety of cell types. A large number of molecules have been identified as modulators of processes required for the enhancement of tumor perfusion. A selected number will be mentioned below. For a recent comprehensive review of molecular and cellular mediators, see Ref. 9.

Vascular endothelial growth factor

Initially named vascular permeability factor, VEGF is a glycoprotein that induces endothelial permeability and func-

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tions as a mitogenic and survival factor of endothelial cells.¹⁰ There are five family members in mammalian cells, VEGF-A to -D and placental growth factor (PlGF). VEGF-A is known also as VEGF, and we will use this term in this review. Five isoforms of VEGF exist (VEGF₁₂₁, VEGF₁₄₅, VEGF₁₆₅, VEGF₁₈₉, and VEGF₂₀₆), differing by their affinity to the extracellular matrix and in their clinical importance.¹¹

VEGF binds Flt-1/VEGFR-1 and Flk-1/VEGFR-2/KDR (kinase domain region), tyrosine kinase receptors, expressed by vascular endothelial cells, and by tumor cells and some epithelial cells. VEGFR-2 is a dominant positive regulator of the vascular system. VEGFR-1 has a higher binding affinity for VEGF than VEGFR-2 but lower kinase activity and no mitogenic response. VEGFR-1 knock-out mice die of overgrowth of endothelial progenitors.¹² In contrast, mice expressing a kinase-dead-VEGFR-1 develop normally, suggesting that this protein is a required negative regulator of the VEGF pathway, acting by sequestering ligand molecules.¹³ However, other models demonstrate VEGFR-1 to be a positive angiogenesis regulator,¹⁴ important in the recruitment of circulating endothelial precursors¹⁵ and of inflammatory cells, which in turn secrete proangiogenic mediators. In addition, VEGFR-1 and VEGFR-2 cross-phosphorylate and activate each other when activated by PlGF and VEGF, respectively. The precise role of VEGFR-1 might be context dependant.

Much less is known about the other members of the VEGF pathway. VEGF-C and VEGF-D mainly regulate lymph vessel formation through activation of VEGFR-3,¹⁶ although VEGFR-3 activity also seems to be required for blood vessel angiogenesis.¹⁷

VEGFR-2 ligand binding causes receptor dimerization, tyrosine phosphorylation, and activation of downstream signaling. Neuropilin-1 is a modulating coreceptor of VEGFR-2 activity. High neuropilin-1 mRNA levels in NSCLC tumors were found to be an independent negative prognostic factor.¹⁸ Another modulator of VEGFR is vascular endothelial-cadherin, which forms a complex with VEGFR-2, β -catenin, and PI3K and is required for the transmission of the survival signal.¹⁹ The VEGF pathway is a major regulator of angiogenesis, cross-talking with other growth factor signaling pathways, some of which are mentioned below.

Platelet-derived growth factor

The platelet-derived growth factor (PDGF) ligands, secreted by tumor cells and endothelial cells, activate PDGF receptors, expressed by pericytes, thus recruiting them to developing vessels. Pericytes have a supportive and modulating role for blood vessels. Abnormal pericyte vessel coverage in PDGF-B null mice results in vessel dilatation, leakage, and hemorrhage.²⁰ The PDGF pathway is also involved in cancer-associated fibroblasts signaling and in autocrine cancer stimulation.⁹ The importance of the PDGF pathway in NSCLC is supported by the correlation found between high tumor PDGF expression levels and poor prognosis.²¹ Preclinical studies indicated enhanced antiangiogenic efficacy of a combined inhibition of VEGFR and PDGFR β .²² Notably, in some cases, platelet-derived endothelial cell growth factor, which is the enzyme thymidine phosphorylase,

produced by the *TYMP* gene, is mistakenly referred to as PDGF.

Angiopoietin (Ang)

The angiopoietin family is composed of four ligands that bind the Tie-2 tyrosine kinase receptor. Ang-1 and Ang-4 function mostly as positive regulators, whereas Ang-2 and Ang-3 are mostly antiangiogenic. However, these roles are context dependent. Ang-2 antagonizes the Ang-1-dependent recruitment of pericytes to new blood vessels, thus preventing their stabilization. Ang-2 knockout mice have defects in adult vascular sprouting, suggesting that destabilization of the vessel structure is required for angiogenesis to progress. Ang-2 seems to have a proangiogenic role when VEGF is abundant but an antiangiogenic role when VEGF levels are low.²³ High tumor Ang-2 expression has negative prognostic implications in lung cancer patients, especially when VEGF expression is high.²⁴ Ang-1 has a negative role in tumor angiogenesis, probably secondary to enhanced pericyte vessel coverage and reduced vessel permeability. The resultant vessels do not allow extravasation of plasma proteins and the microenvironment formed is less proangiogenic.

Endogenous antiangiogenic factors

Potent antiangiogenic factors can be produced endogenously by cleavage of other proteins. For example, angiostatin, a potent angiogenesis inhibitor, is the cleavage product of plasminogen, a component of the coagulation control mechanism. Macrophage-derived methalloelastase (MMP-12) is thought to be responsible for the *in vivo* conversion of plasminogen to angiostatin.²⁵ Thus, tumor-infiltrating macrophages may determine the production of antiangiogenic factors. In addition, cancer cells may secrete enzymes required for the production of angiostatin. Endostatin is an antiangiogenic factor that is a fragment of collagen-XVIII. Additional potent antiangiogenic factors are produced by the cleavage of common proteins,²⁶ suggesting that the tight control of angiogenesis requires reserves of antiangiogenic factors ready for rapid mobilization. Thrombospondin-1 is another endogenous angiogenic inhibitor (AI), mimetics of which are in early clinical trials.²⁷

Cell-cell adhesion molecules

The formation of new blood vessels requires interactions among endothelial cells, pericytes, smooth muscle cells, inflammatory cells, and epithelial cells and involves cell-cell adhesion molecules. Intercellular adhesion molecule 1 (ICAM-1) is a transmembrane protein involved in endothelial cell survival and migration. Plasma ICAM levels were found to be prognostic in a study of lung cancer patients treated with an anti-VEGF antibody.²⁸ N-Cadherin is another cell-cell interaction molecule being evaluated currently as a target for antiangiogenic treatment.²⁹

Growth factors and cytokines

Basic FGF is a potent angiogenic factor that stimulates the proliferation and migration of endothelial cells and the production of matrix metalloproteases (MMPs). Importantly, resistance to VEGFR inhibition can arise from up-regulation of FGF signaling.³⁰ Cytokines also modulate angiogenesis, conceivably by regulating leukocyte recruitment. Interleukin

(IL)-8, IL-12,³¹ and transforming growth factor β are examples of cytokines found to correlate with lung cancer angiogenesis. These and other cytokines are being investigated as predictive factors of efficacy or as therapeutic targets.^{32,33}

Agents Targeting Angiogenesis in Lung Cancer

Targeting the VEGF ligand

Bevacizumab ("Avastin," Roche, Basel, Switzerland) is a monoclonal antibody directed against the VEGF ligand (VEGF-A specific). It was the first bonafide AI to be approved for cancer treatment, initially in colorectal carcinoma. Single agent bevacizumab improved progression-free survival (PFS) in a few trials but seemed inefficient by itself in most cases, and its administration to NSCLC patients in combination with chemotherapy was tested. A phase II study comparing carboplatin and paclitaxel with or without bevacizumab demonstrated improved PFS and a trend of improved overall survival (OS) in the experimental arm³⁴ and led to a phase III trial. Because of cases of fatal pulmonary hemorrhages in squamous cell carcinoma patients in the phase II trial, bevacizumab was henceforth mostly tested in nonsquamous cell lung cancers and is currently approved only for nonsquamous cell lung cancers.

The Eastern Cooperative Oncology Group evaluated bevacizumab in a phase III study of 878 advanced nonsquamous-cell cancer patients (ECOG 4599) randomized to receive paclitaxel and carboplatin with or without bevacizumab. There was a statistically significant increase in median survival from 10.3 to 12.3 months favoring the bevacizumab arm.⁵ This trial led to the approval of bevacizumab for advanced nonsquamous NSCLC as first line regimen. Bevacizumab treatment was also evaluated in combination with cisplatin and gemcitabine (the AVAiL study) in 1050 advanced NSCLC patients and was shown to significantly increase PFS from 6.1 to 6.5 or 6.7 months (in the high and low dose groups, respectively), thus meeting its primary end point. No difference in OS was found in this study.³⁵ The reason for the difference in the results of the AVAiL and the E4599 is not known but could be due to differences in the patient cohorts, the inferior activity of paclitaxel and carboplatin versus cisplatin and gemcitabine, and the *in vitro* data supporting synergy between taxanes and bevacizumab.³⁶ In the BeTa-lung phase III trial, bevacizumab was added to erlotinib ("Tarceva," Roche, Basel, Switzerland) as second-line treatment. PFS was improved from 1.7 to 3.4 months, but only a trend toward survival benefit, the trial's primary end point, was shown.³⁷ Recently, two studies of bevacizumab combined with chemoradiotherapy have reported an alarmingly high rate of tracheoesophageal fistula formation.³⁸ However, another study has reported acceptable toxicity³⁹ and studies are ongoing. As with many other agents that demonstrate activity in the advanced setting, bevacizumab is also being evaluated in the adjuvant setting, in the randomized phase III study ECOG 1505 (NCT00324805). Phase III studies testing AIs are summarized in Table 1 and selected phase I and II studies are presented in Table 2.

VEGF-trap ("Aflibercept," NSC-724770, AVE0005, Regeneron, Inc., Sanofi-Aventis, Paris, France) is a novel

agent designed to function similarly to bevacizumab. VEGF-trap is a recombinant protein, similar to the extracellular-ligand binding domains of VEGFR-1 and 2. It binds all VEGF isoforms and binds VEGF and VEGF-B with a higher affinity than bevacizumab. A phase II study reported some responses in heavily pretreated patients,⁶¹ leading to an ongoing phase III trial, testing the addition of aflibercept to docetaxel as a second-line treatment (NCT00532155).

Targeting the VEGF receptors

Fueled by the success of bevacizumab in advanced NSCLC and by evidence of efficacy of VEGFR inhibitors in other malignancies, much effort have been invested in testing VEGFR inhibitors in NSCLC. Most appealing is the possibility of inhibiting several of the pathways involved in angiogenesis simultaneously. This can be achieved by tyrosine kinase inhibitors (TKIs), which frequently target multiple tyrosine kinase receptors. Most of them co-target the VEGFRs, PDGFRs, and c-Kit, related to their common split-kinase structure. TKIs have the convenience of oral administration. Their short half-life necessitates daily dosing but facilitates the control of reversible toxicities. Bioavailability, pharmacokinetic, and pharmacodynamic properties contribute to differences among the TKIs. Importantly, multikinase inhibition might be antagonistic in some cases. For example, if inhibiting VEGFR-1 would have in tumors a similar effect as knocking it out in mice, enhanced angiogenesis might be expected. Most TKIs, including some claiming high specificity, actually inhibit a significant number of kinases, as demonstrated in an *in vitro* study testing the effect of 38 TKIs on a set of 317 kinases.⁶² Because of the technical challenges inherent in the assessment of kinase inhibition, indirect comparisons of different TKIs, and TKI-defined target claims, should be evaluated cautiously. This group of agents includes most of AI agents tested in clinical trials today (Table 1). Below, we discuss aspects of some of these TKIs.

Sunitinib (SU11248, Pfizer, New York, NY) was developed as an oral inhibitor of VEGFR-1–3, PDGFR- α and β , and c-Kit receptor. More extensive molecular analysis demonstrates it to be one of the most promiscuous TKIs, inhibiting 18% of the 317 tested kinases.⁶² An approved drug for the treatment of gastrointestinal stromal tumor and metastatic renal cell carcinoma (RCCa), it is being tested now as a treatment for NSCLC. Sorafenib (Nexavar, Bayer, Germany) is another TKI, also approved as a treatment for RCCa and for hepatocellular carcinoma. It was designed to target Raf kinase but found to inhibit also VEGFR-1–3, PDGFR, RET, KIT, and FLT-3. A phase III trial comparing chemotherapy with and without sorafenib (ESCAPE trial), closed after it failed to show any benefit with the sorafenib combination.⁶³ Importantly, in the squamous-cell cancer subgroup of patients ($N = 219$), sorafenib-treated patients' survival was reduced from 13.7 to 8.9 months. This is reminiscent of the squamous-cell specific detrimental effect seen with bevacizumab and also noted with Motesanib (AMG706, Amgen, CA) another VEGFR/PDGFR TKI. A detrimental effect of some AI drugs in a subset of patients does not indicate necessarily that this AI or the whole class should be abandoned; rather, it stress the need for biomarkers that will target those drugs to

TABLE 1. Phase III Studies Evaluating Antiangiogenic Treatments

Drug	Tx Line	Treatment	Experimental Arms	Remarks	Name of Study/REF/Clinical Trial, NIH Website
Bevacizumab	I	Carboplatin-paclitaxel	Bevacizumab/placebo	Improved OS (10.3–12.3 mo)	ECOG 4599 ⁵
Bevacizumab	I	Cisplatin-gemcitabine	Bevacizumab/placebo	No OS improvement	AVAIL ^{3,5}
Bevacizumab	II	Erlotinib	Bevacizumab/placebo	Second line Tx. A trend of OS improvement	BeTa-Lung ³⁷
Bevacizumab	Adj	Adj chemotherapy	Bevacizumab/placebo	Various platinum combinations allowed, recruiting	NCT00324805
Ramucirumab	II	Docetaxel	Ramucirumab/placebo	Recruiting	NCT01168973
Aflibercept	II	Docetaxel	Aflibercept/placebo	Ongoing, not recruiting	VITAL, NCT00532155
Sunitinib	II-III	Erlotinib	Sunitinib/placebo	Second or third line treatment, no OS benefit	NCT00457392 ⁴⁰
Sunitinib	Maint	Maint after cisplatin-based chemotherapy	Sunitinib/placebo	Recruiting	NCT00693992
Sorafenib	I	Carboplatin-paclitaxel	Sorafenib/placebo	Stopped at interim analysis: failed, higher mortality in SCCa patients	ESCAPE, NCT00300885
Sorafenib	I	Cisplatin-gemcitabine	Sorafenib/placebo	Squamous cell carcinoma pts withdrawn. No OS or PFS benefit.	NEXUS, NCT00449033 ⁴¹
Cediranib	I	Carboplatin-paclitaxel	Cediranib 30 mg/placebo	Improved RR, higher treatment related mortality	BR.24 ⁴²
Cediranib	I	Carboplatin-paclitaxel	Cediranib 20 mg/placebo	Recruiting	BR.29, NCT00795340
Motesanib (AMG-706)	I	Carboplatin-paclitaxel	Motesanib/placebo	Recruiting. SCCa patients excluded after interim safety review.	MONET1, NCT00460317
Pazopanib	Adj		Pazopanib/placebo	Recruiting. stage I, $T \leq 5$ cm, N0. 24 wk treatment	NCT00775307
Pazopanib	Maint		Pazopanib/placebo	Nonprogressors after first line Tx. Treatment till progression or toxicity	NCT01208064
Vandetanib	II	Docetaxel	Vandetanib/placebo	Increase PFS but not OS, endpoint of study met	Zodiac ^{43,44}
Vandetanib	II	Pemetrexed	Vandetanib/placebo	No improvement	ZEAL ⁴⁵
Vandetanib	II-III	Erlotinib	Vandetanib/Erlotinib	Noninferior	ZEST ⁴⁶
Vandetanib	II-III	Best supportive care	±Vandetanib	EGFR TKI failure patients, OS similar to placebo	Zephyr ⁴⁷
BIBF-1120 (Vargatef)	II	Second line treatment with docetaxel	BIBF-1120/placebo	Recruiting	LUME-lung 1, NCT00805194
BIBF-1120 (Vargatef)	II	Second line treatment with pemetrexed	BIBF-1120/placebo	Recruiting	LUME-lung 2, NCT00806819
Endostar	I	Vinorelbine-cisplatin	Recombinant endostatin/placebo	Improved RR and TTP	Ref. 48
Endostar	Adj	Cisplatin and vinorelbine	±Endostar	Recruiting. stages Ib-IIIa.	NCT00576914 ⁴⁹
Endostar	I	Cisplatin and docetaxel	±Endostar	Recruiting	NCT00657423
ASA404	I	Carboplatin-paclitaxel	ASA404/placebo	Terminated after a negative interim analysis	ATTRACT1, NCT00662597 ⁵⁰
ASA404	II	Docetaxel	ASA404/placebo	Terminated after a negative interim analysis	ATTRACT2, NCT00738387 ⁵¹
Thalidomide	I	Gemcitabine-carboplatin	Thalidomide/placebo (2 yr)	No benefit. Reduced survival in nonsquamous cancer patients	Ref. 52

Tx, treatment; OS, overall survival; PFS, progression free survival; RR, response rate; EGFR, endothelial growth factor receptor; TKI, tyrosine kinase inhibitor; Maint, maintenance; Adj, adjuvant; TTP, time to progression.

TABLE 2. Selected Phase I and II Studies Using AIs

Molecular Mechanism	Name of Drug	Company	Phase I	Phase II	Randomized Phase II
Peptibody: ang inhibitor	CovX	Pfizer, NY	Ongoing ⁵³		
Peptibody: ang inhibitor	AMG 386	Amgen, CA	Ongoing ⁵⁴		
Thrombospondin1 mimic	Cvx-045	Pfizer, NY	Ongoing, not recruiting ²⁷		
Inhibition of VEGF expression	PTC299	PTC therapeutics, NJ	Ongoing ⁵⁵		
Antibody against ALK1	PF-03446962	Pfizer, NY	Ongoing ⁵⁶		
Cyclic peptide: N-Cadherin inhibitor	ADH-1, Exherin	Adherex, Durham, NC	Ongoing, not recruiting NCT00265057		
TKI: VEGFR-1-3, PDGFR- α and β	Tivozanib (AV 951)	Aveo pharmaceuticals	Ongoing, not recruiting (phase I-II) NCT00826878		
Antibody against VEGF	Bevacizumab	Roche, Basel, Switzerland	With chemo-rads (phase I-II) NCT00334815 NCT00280150 NCT00369551	With metronomic chemo NCT00655850 NCT00755170 NCT00755157	Combinations first line, NCT00866528 second line NCT01027598 NCT00871403 NCT01107652
Antibody against VEGF	Bevacizumab	Roche, Basel, Switzerland		With pemetrexed as second line for nonsquamous ⁵⁷ Single agent third line NCT01049776	With chemo NCT01160744 With chemo NCT01160601 NCT01138163 Randomized withdrawal NCT00633789
TKI: VEGFR1-3, PDGFR, and c-Kit receptor	Pazopanib (GW786034)	Glaxo-SmithKline, UK			First line with chemo ⁵⁹ NCT00716534
Antibody against VEGFR-2	Ramucirumab	Eli Lilly, IN			First line, with chemo NCT00768755 NCT00600821
Antibody against phosphatidylserine	Bavituximab	Peregrine, CA			With chemo and bevacizumab NCT00653939
TKI: FGFR and EGFR	Brivanib (BMS-582664)	Bristol-Myers Squibb, NY			
TKI: VEGFR1-3, PDGFR, c-Kit receptor, and FLT3	Linifanib (ABT-869)	Abbott, IL			
TKI: VEGFR1-3, PDGFR, and c-Kit receptor	Axitinib (AG013736)	Pfizer, NY			
Vascular disrupting agent	Fosbretabulin, (CA4P)	OxIGENE, CA			

Peptibody—a hybrid molecule of a targeted peptide and a stabilizing Fc component of an antibody.⁶⁰
TKI, tyrosine kinase inhibitor; VEGFR, vascular endothelial growth factor receptor; Curbo, carboplatin; Chemo-rads, chemotherapy-radiotherapy; Chemo, chemotherapy.

the patients most likely to benefit from it and prevent its use in patients that would be harmed by it. Cediranib (AZD2171, "RECENTIN," AstraZeneca, London, UK), another oral inhibitor of VEGFR-1–3, PDGFR- β , and c-Kit receptor, is an example of this point. A phase II–III trial (BR.24, NCIC-CTG) examined the addition of this agent to a carboplatin and paclitaxel regimen.⁴² Significant toxicities required a reduction of the cediranib dose (from 45 to 30 mg daily). Furthermore, although the death rate was similar in the two arms of the study, a higher rate of treatment-related death was noted in the cediranib arm. However, the response rate increased from 16 to 38% and the hazard ratio for progression-free survival was 0.77 in favor of the cediranib arm⁴² (in the 30 mg cohort, the median progression-free survival was 5.6 months versus 5 months; in the 45 mg cohort, 6.05 versus 5.45 months in the cediranib and placebo arms, respectively). A positive impact of cediranib was seen regardless of gender, histology, or smoking status. Baseline weight loss of more than 5% and hypoalbuminemia predicted increased toxicity for patients receiving cediranib. This led to the initiation of a second trial similar in design, using a lower dose (20 mg) of cediranib, and excluding patients with a weight loss of more than 5% (NCIC-CTG BR.29). Using this approach, the investigators hope to sustain the positive impact of cediranib while reducing toxicity.

An example of a non-TKI that targets the VEGFR is Ramucirumab (Eli Lilly, IN), a novel antibody directed against VEGFR-2, unlike bevacizumab that targets the VEGF ligand.⁵⁸ Directed against the extracellular component of the receptor, its mechanism of inhibition is different than the TKIs, thus in theory both might be applied in concert.

Targeting the EGFR and additional receptors

Vandetanib (ZD6474, "Zactima," AstraZeneca) is an oral inhibitor of VEGFR-2, 3, Ret kinase, and EGFR, conceived with the idea that blocking both angiogenesis and tumor cell proliferation would be synergistic. This drug showed promising activity in advanced NSCLC in several phase II trials, leading to four recently reported phase III studies. None of these trials demonstrated an improvement in survival for patients receiving vandetanib, although the ZODIAC trial met its end point of improved PFS (Table 1). The failure of vandetanib to improve OS might be related to the fact that it inhibits EGFR about 10-fold less than it does VEGFR2⁶⁴ and thus might not actually target both intended pathways. XL 647 (Exelixis, CA) is another oral TKI that targets the VEGFR and EGFR pathways, and inhibits Her2 and EphB4, that was tested in NSCLC patients, demonstrating moderate activity in patients harboring EGFR mutations.⁶⁵ Brivanib (BMS-582664, Bristol-Myers Squibb, NY) is a TKI targeting FGFR and EGFR, thus targeting both angiogenesis and tumor cell proliferation, currently in phase II trials (NCT00633789).

Combining different agents is another manner of inhibiting both the EGFR and the VEGFR pathways. Sorafenib addition to erlotinib treatment did not improve disease-free survival although disease control rate was better (D. Spigel, personal communication, February 2010). Cetuximab with bevacizumab and chemotherapy⁶⁶ is currently being evalu-

ated (NCT00946712), bevacizumab being given to all eligible patients, and randomization done between addition of cetuximab or not. It should be noted that addition of cetuximab to a bevacizumab-chemotherapy regimen worsened outcomes of colorectal cancer patients in two recent trials.^{67,68}

Targeting existing blood vessels (vascular disrupting agents)

Unlike AIs that aim to prevent the sprouting of vessels or the production of new ones, vascular disrupting agents (VDA) target existing blood vessels. VDA can bring about a collapse of the tumor's vascular supply and massive necrosis within hours. Besides the title, various VDA have little in common, embracing a range of mechanisms of action. There are antibodies or peptides that target toxins to tumor-vasculature-specific epitopes, tubulin-binding molecules that target dividing cells and others that are activators of cytokine production. A characteristic observation in studies of VDA is the viable rim of live tumor cells, the culprit of repopulation that remains around an area of central necrosis.⁶⁹ For this reason, VDAs are considered best combined with chemotherapy, radiotherapy, AIs, or other agents that would disrupt this viable rim. Combretastatin A4 phosphate (CA4P, "fosbretabulin," OXiGENE, CA) inhibits microtubule assembly, whereas 5,6-dimethylxanthine-4-acetic acid (DMXAA, ASA404, vadimezan) functions through a different mechanism, partly by activating interferon- β . Originally isolated from the South African willow tree *Combretum caffrum*, these agents block tumor blood flow rapidly by causing vasoconstriction of tumor-feeding arterioles, or by endothelial cell apoptosis.⁷⁰ A promising randomized phase II study with ASA404 has led to two phase III trials, both of them recently reported as negative.^{50,51} CA4P is currently in phase II studies.⁷¹ ABT-751 is a novel antimicrotubule agent also defined as VDA. A phase I–II of this agent in combination with pemetrexed was recently reported with interesting evidence of activity in squamous cell carcinoma patients.⁷²

Targeting angiogenesis using chemotherapy

In *in vivo* models, chemotherapy given at doses much lower than the maximal tolerated dose, but on a continuous basis (metronomic treatment), has a marked antiangiogenic effect.⁷³ Ongoing trials are evaluating this strategy. Some chemotherapy agents targeting the cytoskeleton are potent endothelial cells toxins, e.g., paclitaxel and docetaxel.^{74,75} Furthermore, paclitaxel and docetaxel cause a reduction of interstitial fluid pressure (IFP) by killing tumor cells, thus allowing better tissue perfusion and better drug delivery.⁷⁶

Targeting other angiogenic mechanisms

A recombinant version of endostatin, an endogenous antiangiogenic factor discussed above, was evaluated in 42 patients with neuroendocrine tumors with no documented responses.⁷⁷ Endostar, (YH-16, Simcere, China) is a recombinant human endostatin modified by the addition of nine amino acids, thus simplifying the purification process and improving stability. In a study of 493 advanced NSCLC patients, addition of endostatin to chemotherapy led to increased time to progression, from 3.6 to 6.3 months.⁴⁸ Recently approved in China for NSCLC patients, it is being

tested in various settings including two phase III trials in metastatic disease.

N-Cadherin is a cell-cell adhesion molecule, expressed by endothelial cells and some cancer cells. ADH-1 is an inhibitor of N-Cadherin (Exherin, Adherex, Durham, NC) that was recently tested in phase I–II studies that included NSCLC patients (NCT00265057, NCT00264433). Thalidomide has an antiangiogenic activity that is poorly understood and may act indirectly through its immunomodulatory effects. A negative phase III study testing its role in NSCLC was recently published.⁵²

Targeting tumor vasculature-specific proteins is an additional approach which seems appealing as a therapeutic strategy. Tumor-blood-vessel-specific proteins and phospholipids have been identified. One of these is phosphatidylserine, normally found only in the inner lipid leaflet of the cell membrane but translocates to the outer leaflet in tumor vasculature. It is currently targeted in the clinic by a novel specific antibody (bavituximab, Peregrine, CA).⁷⁸

Insight from Studies Using AIs in Lung Cancer

Resistant angiogenesis

Although many AIs show promising responses and PFS improvements in NSCLC patients, almost none bring about prolongation of survival. The reasons for this phenomenon are not known. A relevant observation might be rebound angiogenesis, observed in mice studies where the VEGF pathway was effectively attenuated.³⁰ In these models, alternative angiogenic mechanisms are up-regulated, including induction of FGF family members, angiopoietins, and vessel co-option, possibly triggered by hypoxia in tumors subjected to VEGF inhibition.⁷⁹ Indeed, up-regulation of one of several alternative angiogenic pathways has been demonstrated in human lung cancers.⁸⁰ Another important finding in models of VEGF-dependent tumor growth treated with VEGF inhibitors is increased invasiveness of tumors that thrive in these conditions.^{30,81} This could be secondary to hypoxia-induced activation of the hepatocyte growth factor-Met pathway,⁸² urokinase-type plasminogen activator,⁸³ or other survival pathways. Furthermore, hypoxia may select for tumor cells with an aggressiveness phenotype, e.g., those with loss of p53⁸⁴ or other genetic events.⁸⁵ Two recent studies in mice models indicate a risk of enhanced metastatic spread as a result of AI treatment.^{86,87} Using VEGF pathway inhibition as a cancer therapeutic strategy, requires further understanding about cancer escape and resistance mechanisms.

The requirement for a continuous treatment

The importance of treatment schedules of AIs is currently not clear. Rebound accelerated tumor growth is seen on discontinuation of bevacizumab treatment of metastatic colorectal cancer⁸⁸ and in patients with glioblastoma multiforme during drug holiday of a VEGFR inhibitor.⁸⁹ Sunitinib and axitinib treatment breaks result in increase tumor perfusion and proliferation.⁹⁰ In a retrospective comparison of two cohorts of a sunitinib phase II study, more responses were seen in a noncontinuous, higher dose treatment, but longer survival was observed in the cohort treated continuously,⁹¹ supporting the principle of daily administration of AI drugs.

Regarding the length of treatment, current practice with most AIs is continuous treatment till progression. Supportive of this, maintenance bevacizumab prolonged PFS in a recent phase III trial of ovarian, peritoneal, and fallopian carcinomas.⁹² When combined with chemotherapy, it is not known whether AIs should be continued beyond progression. We are not aware of any studies designed to answer this question.

Combining AI treatments with chemotherapy

The only trial where an AI prolonged the life of NSCLC patients involved administering bevacizumab with chemotherapy. However, when an AI agent is given concurrently with a cytotoxic agent, antagonism is expected, because damaging the tumor's blood supply should hinder delivery of the cytotoxic agents. A possible explanation for the apparently unexpected synergism observed clinically was suggested by Jain and others regarding the IFP of tumors.⁹³ The increased permeability typical of tumor vessels results in extravasation of macromolecules and fluid to the extravascular compartment and increased IFP. This increased IFP reduces vascular flow and drug delivery. AIs cause a rapid normalization of tumor vasculature, reducing their permeability and the IFP. Thus, before elimination of the vessels that perfuse a tumor, a window of opportunity might exist when tumor perfusion and drug delivery is increased.⁹⁴ IFP is reduced after treatment with anti-VEGF antibody,⁹³ with a VDA,⁹⁵ and with sunitinib,⁹⁶ cediranib⁸⁹ and pazopanib (Votrient, GlaxoSmithKlin, UK).⁹⁷ However, improved tumor perfusion after such treatments remains a theoretical consideration and to date has not been consistently demonstrated by perfusion studies in humans. In contrast, axitinib decreased tumor exposure to concomitantly given cyclophosphamide,⁹⁸ and sunitinib treatment only enhanced day 3 diffusion parameters and not tumor perfusion.⁹⁹ On the basis of those observations, it can be speculated that an AI-drug holiday before chemotherapy administration might enhance chemotherapy delivery and treatment efficacy. However, AI and chemotherapy synergism might be secondary to enhanced killing of tumor endothelial cells,^{4,75,98} suggesting they should be given concurrently. The requirement for an AI-drug holiday is being investigated in a phase I–II trial combining axitinib with cisplatin/pemetrexed (NCT00768755).

Taxanes were shown to mobilize bone marrow-derived circulating endothelial progenitor cells (CEPs) that colonize tumors and allow for regeneration of tumor vasculature after treatment.³⁶ Blocking the VEGF pathway prevents the CEPs surge and might be basis for chemotherapy-AI synergism, possibly limited to drugs that mobilize CEPs.

Evaluating response to AI treatments

An apparent class effect of AI treatments is tumor cavitation, seen in 24% of treated tumors in one published series.¹⁰⁰ It seems plausible that assessing response solely according to RECIST criteria might misclassify cavitating tumors. An alternative single dimension measurement taking cavitation size into account was recently proposed.¹⁰⁰ Software-assisted volumetric measurement is an alternative approach. The role of these novel response measurements needs to be examined in large trials, comparing them to standard RECIST criteria.

Toxicities of AIs

Less than 1% of adult endothelial cells are actively proliferating, suggesting that targeting endothelial proliferation would impact only tumor-activated endothelial cells and wound healing and be devoid of significant side effects. However, the toxicities seen with VEGFR inhibitors indicate an unappreciated role of VEGF in microvessels and tissue homeostasis. Further discussion of this topic is beyond the scope of this review. Interestingly, some toxicities might be useful as predictive markers of response to treatment.¹⁰¹

Predicting Response to AI Treatments

In light of the potentially significant toxicities and lack of survival benefit in many of the AI trials, there is an urgent need for predictive biomarkers. In an analysis of the E4599 trial, VEGF plasma levels were predictive of response to bevacizumab but not predictive of a survival benefit.²⁸ In contrast, VEGF plasma levels were negatively correlated with PFS in patients treated with vandetanib,¹⁰² and a greater increase in VEGF plasma levels with vandetanib treatment predicted worse outcome.³² PIGF elevation showed a trend to be predictive of response to motesanib.¹⁰³ Polymorphisms in the *VEGF* gene, and in the *ICAM-1* and *WNK1* genes were found to correlate with bevacizumab-related improved survival.¹⁰⁴ An increase in ICAM-1 levels with treatment was associated with a better PFS in vandetanib-treated NSCLC patients.³² Analysis of E4599 samples revealed improved PFS with bevacizumab for patients with low baseline levels of ICAM-1 and improved OS with bevacizumab for patients with stable levels of E-selectin.²⁸ Baseline levels of hepatocyte growth factor and of IL-12 were predictive of response to pazopanib.³³ Tumor mRNA levels of LDH-A, Glut-1, and VEGFR-1 were found to be predictive of response to the VEGFR inhibitor PTK787/ZK 222584 in colorectal cancer patients.¹⁰⁵

In vivo measurements of patients' tumor blood perfusion changes with initial doses of the antiangiogenic agent are potential predictive markers. Magnetic resonance imaging dynamic contrast enhancement measurements after 2 days of an oral antiangiogenic treatment were found to correlate with patients' drug exposure.¹⁰⁶ As early as 24 hours after the first oral dose of an angiogenesis inhibitor, a significant reduction of permeability and vessel size can be demonstrated.⁸⁹ The predictive power of these imaging studies is yet to be validated.

Clinical characteristics might also be predictive. Female patients did not benefit from bevacizumab in the E4599,¹⁰⁷ whereas an opposite trend was seen in the AVaIL trial³⁵ and in a phase II vandetanib trial³² (a result not reproduced in the phase III vandetanib trial⁴³). Adenocarcinoma patients had a clear benefit from bevacizumab, a result that could not be demonstrated for other nonsquamous histologies.¹⁰⁸ Recently, antiangiogenesis-induced arterial hypertension was found to correlate with clinical response and outcome in various tumors.^{109–111} Retrospective analysis of the E4599 NSCLC trial demonstrates that HTN induced by bevacizumab correlates with a significantly improved OS¹⁰¹ but conflicting results have been reported in colorectal cancer.¹¹² So far, none of the above

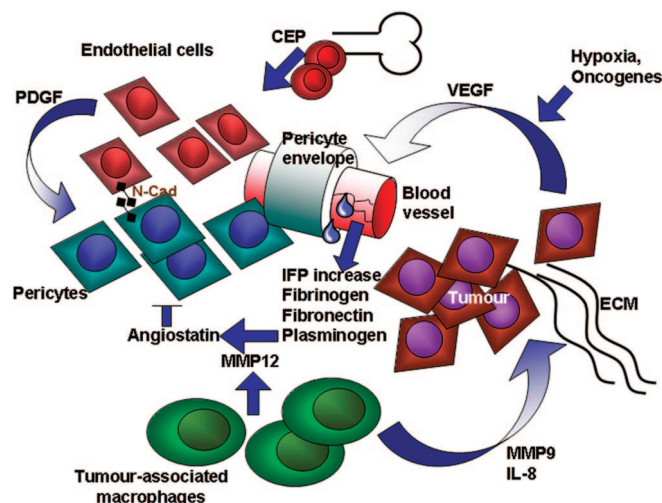


FIGURE 1. Vascular endothelial growth factor (VEGF) secretion by tumor cells is induced by tumor hypoxia and oncogene signaling.⁴ VEGF acts mainly on endothelial cells, inducing angiogenesis and a hyperpermeable state. Tumor endothelial cells are one of the sources of platelet-derived growth factor, which recruit pericytes to the vessels' envelope. N-Cadherin (N-Cad) physically connects endothelial cells and pericytes.²⁹ Plasma proteins such as fibrinogen and fibronectin leak through the abnormal tumor vessels, contributing to the proangiogenic microenvironment and to the enhanced interstitial fluid pressure (IFP) typical of tumors.⁹⁴ Tumor-associated macrophages are a subtype of macrophages that invade cancer stroma and secrete MMP9, leading to release of extracellular matrix-associated VEGF¹¹³; these cells are also capable of promoting the formation of angiogenic factors such as angiostatin.²⁵ Circulating endothelial progenitors (CEP) are putative bone marrow-derived cells that home to tumor sites and might be a major contributor to the newly formed blood vessels.⁶⁹ This scheme represents a partial view of the major players involved in cancer angiogenesis.

mentioned candidate predictive biomarkers have been validated prospectively.

Summary and Future Directions

Angiogenesis inhibitors have obvious antitumor activity in NSCLC and bevacizumab has become part of the standard of care for patients with advanced nonsquamous cell carcinoma. Unfortunately, to date, the gains have been modest. As we learn more about angiogenesis, the complexity of the biology becomes apparent (Figure 1) and the number of unanswered questions increase (Table 3). As seen in many areas of medicine in the past, some of the answers may be surprisingly simple. New and exciting areas of research include elucidating the role of CEPs, tumor-vasculature specific molecules that allow their specific targeting, novel signaling pathways involved in angiogenesis and in vivo real-life imaging of the vasculature and tumor cell proliferation. These developments and others are eagerly awaited. In conclusion, the tumor vasculature remains an important area of anticancer research. Better understanding of the biology

TABLE 3. Nonresolved Issues Regarding AI for the Treatment of Lung Cancer

Nonresolved issues regarding combination treatments

Blocking multiple angiogenic factors (e.g., VEGFRs and PDGFRs) vs. blocking specific molecular mediators proven to be positive angiogenic regulators (e.g., blocking VEGFR-2 and not VEGFR-1)?

Blocking simultaneously the VEGF signaling pathway (targeting endothelial cells, achieving microvasculature normalization) and the PDGF pathway (targeting pericytes, preventing vessel maturation)?

Blocking simultaneously the extracellular component and internal domain of a relevant signaling pathway (e.g., bevacizumab and VEGFR TKI) to achieve complete blockage?

Blocking angiogenesis and proliferation pathways simultaneously (e.g., vandetanib), thus targeting both tumor cells and their blood supply?

Nonresolved issues regarding the preferred type of therapeutic molecule
Prefer small molecule TKIs

To allow multikinase targeting?

To allow rapid reversal of toxicities (short half-life)?

Prefer humanized antibodies/peptibodies

To allow specific single molecule targeting?

To enhance preferential tumor delivery (through enhanced permeability and retention effect¹¹⁴)?

Nonresolved issues about combining AI with chemotherapy

Combine AI with chemotherapy

As a way to prevent CEP-mediated tumor salvage?

As a way to reduce IFP and improve chemotherapy delivery?

As a way to kill endothelial cells more efficiently?

The need for an AI drug holiday proceeding chemotherapy administration?

Continue AI agents after AI-chemo combined treatment, until disease progression?

Continue AI treatment beyond progression on AI-chemo combined treatment?

AI, angiogenesis inhibitor; chemo, chemotherapy; TKI, tyrosine kinase inhibitor; VEGF, vascular endothelial growth factor; IFP, interstitial fluid pressure; CEP, circulating endothelial progenitors.

and trials with angiogenesis inhibitors has already yielded positive results for NSCLC patients. Ongoing study of this exciting target is imperative.

REFERENCES

1. Folkman J. Tumor angiogenesis: therapeutic implications. *N Engl J Med* 1971;285:1182–1186.
2. Herbst RS, Onn A, Sandler A. Angiogenesis and lung cancer: prognostic and therapeutic implications. *J Clin Oncol* 2005;23:3243–3256.
3. Fontanini G, Lucchi M, Vignati S, et al. Angiogenesis as a prognostic indicator of survival in non-small-cell lung carcinoma: a prospective study. *J Natl Cancer Inst* 1997;89:881–886.
4. Bar J, Onn A, Herbst RS. Molecular events surrounding the angiogenic switch of lung cancer. In HI Pass, DP Carbone, DH Johnson, JD Minna, GV Scagliotti, AT Turrisi III, (Eds.) *Principles and Practice of Lung Cancer*. Philadelphia, PA: Lippincott Williams & Wilkins, 2010. Pp. 113–134.
5. Sandler A, Gray R, Perry MC, et al. Paclitaxel-carboplatin alone or with bevacizumab for non-small-cell lung cancer. *N Engl J Med* 2006;355:2542–2550.
6. Renyi-Vamos F, Tovari J, Fillinger J, et al. Lymphangiogenesis correlates with lymph node metastasis, prognosis, and angiogenic phenotype in human non-small cell lung cancer. *Clin Cancer Res* 2005;11:7344–7353.
7. Pezzella F, Pastorino U, Tagliabue E, et al. Non-small-cell lung carcinoma tumor growth without morphological evidence of neo-angiogenesis. *Am J Pathol* 1997;151:1417–1423.
8. Mattern J, Koomagi R, Volm M. Biological characterization of subgroups of squamous cell lung carcinomas. *Clin Cancer Res* 1999;5:1459–1463.
9. Chung AS, Lee J, Ferrara N. Targeting the tumour vasculature: insights from physiological angiogenesis. *Nat Rev Cancer* 2010;10:505–514.
10. Ferrara N, Gerber HP, LeCouter J. The biology of VEGF and its receptors. *Nat Med* 2003;9:669–676.
11. Yuan A, Yu CJ, Kuo SH, et al. Vascular endothelial growth factor 189 mRNA isoform expression specifically correlates with tumor angiogenesis, patient survival, and postoperative relapse in non-small-cell lung cancer. *J Clin Oncol* 2001;19:432–441.
12. Fong GH, Zhang L, Bryce DM, et al. Increased hemangioblast commitment, not vascular disorganization, is the primary defect in flt-1 knock-out mice. *Development* 1999;126:3015–3025.
13. Hiratsuka S, Minowa O, Kuno J, et al. Flt-1 lacking the tyrosine kinase domain is sufficient for normal development and angiogenesis in mice. *Proc Natl Acad Sci USA* 1998;95:9349–9354.
14. Fischer C, Jonckx B, Mazzone M, et al. Anti-PlGF inhibits growth of VEGF(R)-inhibitor-resistant tumors without affecting healthy vessels. *Cell* 2007;131:463–475.
15. Lyden D, Hattori K, Dias S, et al. Impaired recruitment of bone-marrow-derived endothelial and hematopoietic precursor cells blocks tumor angiogenesis and growth. *Nat Med* 2001;7:1194–1201.
16. Alitalo K, Tammela T, Petrova TV. Lymphangiogenesis in development and human disease. *Nature* 2005;438:946–953.
17. Laakkonen P, Waltari M, Holopainen T, et al. Vascular endothelial growth factor receptor 3 is involved in tumor angiogenesis and growth. *Cancer Res* 2007;67:593–599.
18. Hong TM, Chen YL, Wu YY, et al. Targeting neuropilin 1 as an antitumor strategy in lung cancer. *Clin Cancer Res* 2007;13:4759–4768.
19. Carmeliet P, Lampugnani MG, Moons L, et al. Targeted deficiency or cytosolic truncation of the VE-cadherin gene in mice impairs VEGF-mediated endothelial survival and angiogenesis. *Cell* 1999;98:147–157.
20. Lindahl P, Johansson BR, Leveen P, et al. Pericyte loss and microaneurysm formation in PDGF-B-deficient mice. *Science* 1997;277:242–245.
21. Donnem T, Al-Saad S, Al-Shibli K, et al. Prognostic impact of platelet-derived growth factors in non-small cell lung cancer tumor and stromal cells. *J Thorac Oncol* 2008;3:963–970.
22. Bergers G, Song S, Meyer-Morse N, et al. Benefits of targeting both pericytes and endothelial cells in the tumor vasculature with kinase inhibitors. *J Clin Invest* 2003;111:1287–1295.
23. Lobov IB, Brooks PC, Lang RA. Angiopoietin-2 displays VEGF-dependent modulation of capillary structure and endothelial cell survival in vivo. *Proc Natl Acad Sci USA* 2002;99:11205–11210.
24. Tanaka F, Ishikawa S, Yanagihara K, et al. Expression of angiopoietins and its clinical significance in non-small cell lung cancer. *Cancer Res* 2002;62:7124–7129.
25. Dong Z, Kumar R, Yang X, et al. Macrophage-derived metalloelastase is responsible for the generation of angiostatin in Lewis lung carcinoma. *Cell* 1997;88:801–810.
26. Petitclerc E, Boutaud A, Prestayko A, et al. New functions for non-collagenous domains of human collagen type IV. Novel integrin ligands inhibiting angiogenesis and tumor growth in vivo. *J Biol Chem* 2000;275:8051–8061.
27. Mendelson DS, Dinolfo M, Cohen RB, et al. First-in-human dose escalation safety and pharmacokinetic (PK) trial of a novel intravenous (IV) thrombospondin-1 (TSP-1) mimetic humanized monoclonal CovX Body (CVX-045) in patients (pts) with advanced solid tumors. *J Clin Oncol* 2008;26:abstract 3524 (ASCO Meeting Abstracts).
28. Dowlati A, Gray R, Sandler AB, et al. Cell adhesion molecules, vascular endothelial growth factor, and basic fibroblast growth factor in patients with non-small cell lung cancer treated with chemotherapy with or without bevacizumab—an Eastern Cooperative Oncology Group Study. *Clin Cancer Res* 2008;14:1407–1412.
29. Perotti A, Sessa C, Mancuso A, et al. Clinical and pharmacological phase I evaluation of Exherin (ADH-1), a selective anti-N-cadherin

- peptide in patients with N-cadherin-expressing solid tumours. *Ann Oncol* 2009;20:741–745.
30. Casanovas O, Hicklin DJ, Bergers G, et al. Drug resistance by evasion of antiangiogenic targeting of VEGF signaling in late-stage pancreatic islet tumors. *Cancer Cell* 2005;8:299–309.
31. Tran HT, Kim ES, Lee JJ, et al. Correlation between plasma cytokine/angiogenic factors (C/AF) and signaling pathways activation from baseline tumor biopsy specimens in patients with advanced non small cell lung cancer (NSCLC): preliminary analysis from the Biomarker-based Approaches of Targeted Therapy for Lung Cancer Elimination (BATTLE) Clinical Study. *J Clin Oncol* 2008;26:abstract 8010 (ASCO Meeting Abstracts).
32. Hanrahan EO, Lin HY, Kim ES, et al. Distinct patterns of cytokine and angiogenic factor modulation and markers of benefit for vandetanib and/or chemotherapy in patients with non-small-cell lung cancer. *J Clin Oncol* 2010;28:193–201.
33. Nikolinakos PG, Altorki N, Yankelevitz D, et al. Plasma cytokine and angiogenic factor profiling identifies markers associated with tumor shrinkage in early-stage non-small cell lung cancer patients treated with pazopanib. *Cancer Res* 2010;70:2171–2179.
34. Johnson DH, Fehrenbacher L, Novotny WF, et al. Randomized phase II trial comparing bevacizumab plus carboplatin and paclitaxel with carboplatin and paclitaxel alone in previously untreated locally advanced or metastatic non-small-cell lung cancer. *J Clin Oncol* 2004;22:2184–2191.
35. Reck M, von Pawel J, Zatloukal P, et al. Phase III trial of cisplatin plus gemcitabine with either placebo or bevacizumab as first-line therapy for nonsquamous non-small-cell lung cancer: AVAiL. *J Clin Oncol* 2009;27:1227–1234.
36. Shaked Y, Henke E, Roodhart JM, et al. Rapid chemotherapy-induced acute endothelial progenitor cell mobilization: implications for antiangiogenic drugs as chemosensitizing agents. *Cancer Cell* 2008;14:263–273.
37. Hainsworth J, Herbst RS. A phase III, multicenter, placebo-controlled, double-blind, randomized clinical trial to evaluate the efficacy of bevacizumab (Avastin®) in combination with erlotinib (Tarceva®) compared with erlotinib alone for treatment of advanced non-small cell lung cancer after failure of standard first-line chemotherapy (BETA). *J Thorac Oncol* 2008;3:S302.
38. Spigel DR, Hainsworth JD, Yardley DA, et al. Tracheoesophageal fistula formation in patients with lung cancer treated with chemoradiation and bevacizumab. *J Clin Oncol* 2010;28:43–48.
39. Socinski MA, Stinchcombe TE, Halle JS, et al. Incorporation of bevacizumab (B) and erlotinib (Er) with induction (Ind) and concurrent (Conc) carboplatin (Cb)/paclitaxel (P) and 74 Gy of thoracic radiotherapy in stage III non-small cell lung cancer (NSCLC). *J Clin Oncol* 2009;27:abstract 7528 (ASCO Meeting Abstracts).
40. Scagliotti GV, Krzakowski M, Szczesna A, et al. Sunitinib (SU) in combination with Erlotinib (E) for the treatment of advanced/metastatic Non Small Cell Lung Cancer (NSCLC): a phase III study. Presented at the 35th European Society for Medical Oncology Congress 2010, Milan, Italy, October 8–12, 2010:LBA6.
41. Gatzemeier U, Eisen T, Santoro A, et al. Sorafenib (S) + gemcitabine/cisplatin (GC) vs GC alone in the first-line treatment of advanced non-small cell lung cancer (NSCLC): phase III NSCLC research experience utilizing sorafenib (NEXUS) trial. Presented at the 35th European Society for Medical Oncology Congress 2010, Milan, Italy, October 8–12, 2010:LBA16.
42. Goss GD, Arnold A, Shepherd FA, et al. Randomized, double-blind trial of carboplatin and paclitaxel with either daily oral cediranib or placebo in advanced non-small-cell lung cancer: NCIC Clinical Trials Group BR24 Study. *J Clin Oncol* 2009;28:49–55.
43. Herbst RS, Sun Y, Eberhardt WE, et al. Vandetanib plus docetaxel versus docetaxel as second-line treatment for patients with advanced non-small-cell lung cancer (ZODIAC): a double-blind, randomised, phase 3 trial. *Lancet Oncol* 2010;11:619–626.
44. Herbst RS, Sun Y, Korf S, et al. Vandetanib plus docetaxel versus docetaxel as second-line treatment for patients with advanced non-small cell lung cancer (NSCLC): a randomized, double-blind phase III trial (ZODIAC). *J Clin Oncol* 2009;27:abstract CRA8003 (ASCO Meeting Abstracts).
45. de Boer RH, Arrieta O, Yang CH, et al. Vandetanib plus pemetrexed for the second-line treatment of advanced non-small-cell lung cancer: a randomized, double-blind phase III trial. *J Clin Oncol* 2011;29:1067–1074.
46. Natale RB, Thongprasert S, Greco FA, et al. Phase III trial of vandetanib compared with erlotinib in patients with previously treated advanced non-small-cell lung cancer. *J Clin Oncol* 2011;29:1059–1066.
47. Lee J, Hirsh V, Park K, et al. Vandetanib versus placebo in patients with advanced non-small cell lung cancer (NSCLC) after prior therapy with an EGFR tyrosine kinase inhibitor (TKI): a randomized, double-blind phase III trial (ZEPHYR). *J Clin Oncol* 2010;28:abstract 7525 (ASCO Meeting Abstracts).
48. Sun Y, Wang J, Liu Y, et al. Results of phase III trial of rh-endostatin (YH-16) in advanced non-small cell lung cancer (NSCLC) patients. *J Clin Oncol* 2005;23:abstract 7138 (ASCO Meeting Abstracts).
49. Jie H, Xiangru Z, Han B, et al. A phase III adjuvant vinorelbine plus cisplatin (NP) versus NP plus endostar (NPE) in patients (pts) with completely resected stage IB-IIIA non-small cell lung cancer (NSCLC): an interim preliminary result. *J Clin Oncol* 2010;28:abstract 7019 (ASCO Meeting Abstracts).
50. Lara PN Jr, Douillard JY, Nakagawa K, et al. Randomized phase III placebo-controlled trial of carboplatin and paclitaxel with or without the vascular disrupting agent vandimezan (ASA404) in advanced non-small-cell lung cancer. *J Clin Oncol* 2011;29:2965–2971.
51. ATTRACT. ATTRACT website; interim analysis of ATTRACT-2 trial. Available at: <http://www.attractstudy.com/attract-2-researching-ASA404-in-cancer-treatment.jsp>.
52. Lee SM, Rudd R, Woll PJ, et al. Randomized double-blind placebo-controlled trial of thalidomide in combination with gemcitabine and carboplatin in advanced non-small-cell lung cancer. *J Clin Oncol* 2009;27:5248–5254.
53. Rosen LS, Mendelson DS, Cohen RB, et al. First-in-human dose-escalation safety and PK trial of a novel intravenous humanized monoclonal CovX body inhibiting angiopoietin 2. *J Clin Oncol* 2010;28:abstract 2524 (ASCO Meeting Abstracts).
54. Herbst RS, Hong D, Chap L, et al. Safety, pharmacokinetics, and antitumor activity of AMG 386, a selective angiopoietin inhibitor, in adult patients with advanced solid tumors. *J Clin Oncol* 2009;27:3557–3565.
55. Schwartz GK, Dickson MA, Callahan LA, et al. Phase Ib study of PTC299, a novel oral inhibitor of tumor VEGF expression, in patients with advanced cancer. *J Clin Oncol* 2010;28:abstract 3041 (ASCO Meeting Abstracts).
56. Goff LW, De Braud FG, Cohen RB, et al. Phase I study of PF-03449662, a fully human mab against ALK 1, a TGF (beta) receptor involved in tumor angiogenesis. *J Clin Oncol* 2010;28:abstract 3034 (ASCO Meeting Abstracts).
57. Adjei AA, Mandrekar SJ, Dy GK, et al. Phase II trial of pemetrexed plus bevacizumab for second-line therapy of patients with advanced non-small-cell lung cancer: NCCTG and SWOG study N0426. *J Clin Oncol* 2010;28:614–619.
58. Camidge DR, Ballas MS, Dubey S, et al. A phase II, open-label study of ramucirumab (IMC-1121B), an IgG1 fully human monoclonal antibody (MAb) targeting VEGFR-2, in combination with paclitaxel and carboplatin as first-line therapy in patients (pts) with stage IIb/IV non-small cell lung cancer (NSCLC). *J Clin Oncol* 2010;28:abstract 7588 (ASCO Meeting Abstracts).
59. Tan EH, Goss GD, Salgia R, et al. Phase 2 trial of Linifanib (ABT-869) in patients with advanced non-small cell lung cancer. *J Thorac Oncol* 2011;6:1418–1425.
60. Sutherland S. Peptibodies: the new cool technology. *Drug Discov Today* 2004;9:683.
61. Leigh NB, Raez LE, Besse B, et al. A multicenter, phase 2 study of vascular endothelial growth factor trap (Aflibercept) in platinum- and erlotinib-resistant adenocarcinoma of the lung. *J Thorac Oncol* 2010;5:1054–1059.
62. Karaman MW, Herrgard S, Treiber DK, et al. A quantitative analysis of kinase inhibitor selectivity. *Nat Biotechnol* 2008;26:127–132.
63. Scagliotti G, Novello S, von Pawel J, et al. Phase III study of carboplatin and paclitaxel alone or with sorafenib in advanced non-small-cell lung cancer. *J Clin Oncol* 2010;28:1835–1842.

64. Morabito A, Piccirillo MC, Falasconi F, et al. Vandetanib (ZD6474), a dual inhibitor of vascular endothelial growth factor receptor (VEGFR) and epidermal growth factor receptor (EGFR) tyrosine kinases: current status and future directions. *Oncologist* 2009;14:378–390.
65. Haymach J. In Proceedings of the 11th Annual Targeted Therapies of Lung Cancer Meeting, Santa Monica, CA, February 2011.
66. Gandara D, Kim ES, Herbst RS, et al. S0536: Carboplatin, paclitaxel, cetuximab, and bevacizumab followed by cetuximab and bevacizumab maintenance in advanced non-small cell lung cancer (NSCLC): a SWOG phase II study. *J Clin Oncol* 2009;27:abstract 8015 (ASCO Meeting Abstracts).
67. Tol J, Koopman M, Cats A, et al. Chemotherapy, bevacizumab, and cetuximab in metastatic colorectal cancer. *N Engl J Med* 2009;360:563–572.
68. Hecht JR, Mitchell E, Chidiac T, et al. A randomized phase IIIB trial of chemotherapy, bevacizumab, and panitumumab compared with chemotherapy and bevacizumab alone for metastatic colorectal cancer. *J Clin Oncol* 2009;27:672–680.
69. Shaked Y, Ciarrocchi A, Franco M, et al. Therapy-induced acute recruitment of circulating endothelial progenitor cells to tumors. *Science* 2006;313:1785–1787.
70. Sheng Y, Hua J, Pinney KG, et al. Combretastatin family member OXI4503 induces tumor vascular collapse through the induction of endothelial apoptosis. *Int J Cancer* 2004;111:604–610.
71. Garon EB, Kabbinnar FF, Neidhart JA, et al. Randomized phase II trial of a tumor vascular disrupting agent fosbretabulin tromethamine (CA4P) with carboplatin (C), paclitaxel (P), and bevacizumab (B) in stage IIIB/IV nonsquamous non-small cell lung cancer (NSCLC): The FALCON trial. *J Clin Oncol* 2010;28:abstract 7587 (ASCO Meeting Abstracts).
72. Rudin CM, Mauer A, Smakal M, et al. Phase I/II study of pemetrexed with or without ABT-751 in advanced or metastatic non-small-cell lung cancer. *J Clin Oncol* 2011;29:1075–1082.
73. Browder T, Butterfield CE, Kraling BM, et al. Antiangiogenic scheduling of chemotherapy improves efficacy against experimental drug-resistant cancer. *Cancer Res* 2000;60:1878–1886.
74. Belotti D, Vergani V, Drudis T, et al. The microtubule-affecting drug paclitaxel has antiangiogenic activity. *Clin Cancer Res* 1996;2:1843–1849.
75. Sweeney CJ, Miller KD, Sissons SE, et al. The antiangiogenic property of docetaxel is synergistic with a recombinant humanized monoclonal antibody against vascular endothelial growth factor or 2-methoxyestradiol but antagonized by endothelial growth factors. *Cancer Res* 2001;61:3369–3372.
76. Griffon-Etienne G, Boucher Y, Brekken C, et al. Taxane-induced apoptosis decompresses blood vessels and lowers interstitial fluid pressure in solid tumors: clinical implications. *Cancer Res* 1999;59:3776–3782.
77. Kulke MH, Bergsland EK, Ryan DP, et al. Phase II study of recombinant human endostatin in patients with advanced neuroendocrine tumors. *J Clin Oncol* 2006;24:3555–3561.
78. Digumarti R, Suresh AV, Bhattacharyya GS, et al. Phase II study of bavituximab plus paclitaxel and carboplatin in untreated locally advanced or metastatic non-small cell lung cancer: interim results. *J Clin Oncol* 2010;28:abstract 7589 (ASCO Meeting Abstracts).
79. Folkman J. Angiogenesis: an organizing principle for drug discovery? *Nat Rev Drug Discov* 2007;6:273–286.
80. McClelland MR, Carskadon SL, Zhao L, et al. Diversity of the angiogenic phenotype in non-small cell lung cancer. *Am J Respir Cell Mol Biol* 2007;36:343–350.
81. Bergers G, Brekken R, McMahon G, et al. Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol* 2000;2:737–744.
82. Pennacchietti S, Michieli P, Galluzzo M, et al. Hypoxia promotes invasive growth by transcriptional activation of the met protooncogene. *Cancer Cell* 2003;3:347–361.
83. Rofstad EK, Rasmussen H, Galappathi K, et al. Hypoxia promotes lymph node metastasis in human melanoma xenografts by up-regulating the urokinase-type plasminogen activator receptor. *Cancer Res* 2002;62:1847–1853.
84. Graeber TG, Osmanian C, Jacks T, et al. Hypoxia-mediated selection of cells with diminished apoptotic potential in solid tumours. *Nature* 1996;379:88–91.
85. Steeg PS. Angiogenesis inhibitors: motivators of metastasis? *Nat Med* 2003;9:822–823.
86. Ebos JM, Lee CR, Cruz-Munoz W, et al. Accelerated metastasis after short-term treatment with a potent inhibitor of tumor angiogenesis. *Cancer Cell* 2009;15:232–239.
87. Paez-Ribes M, Allen E, Hudock J, et al. Antiangiogenic therapy elicits malignant progression of tumors to increased local invasion and distant metastasis. *Cancer Cell* 2009;15:220–231.
88. Cacheux W, Boissier T, Staudacher L, et al. Reversible tumor growth acceleration following bevacizumab interruption in metastatic colorectal cancer patients scheduled for surgery. *Ann Oncol* 2008;19:1659–1661.
89. Batchelor TT, Sorensen AG, di Tomaso E, et al. AZD2171, a pan-VEGF receptor tyrosine kinase inhibitor, normalizes tumor vasculature and alleviates edema in glioblastoma patients. *Cancer Cell* 2007;11:83–95.
90. Jeraj R, Liu G, Simoncic U, et al. Concurrent assessment of vasculature and proliferative pharmacodynamics in patients treated with VEGFR TKI. *J Clin Oncol* 2010;28:abstract 3050 (ASCO Meeting Abstracts).
91. Socinski MA. The current status and evolving role of sunitinib in non-small cell lung cancer. *J Thorac Oncol* 2008;3:S119–S123.
92. Burger RA, Brady MF, Bookman MA, et al. Phase III trial of bevacizumab (BEV) in the primary treatment of advanced epithelial ovarian cancer (EOC), primary peritoneal cancer (PPC), or fallopian tube cancer (FTC): a Gynecologic Oncology Group study. *J Clin Oncol* 2010;28:abstract LBA1 (ASCO Meeting Abstracts).
93. Willett CG, Boucher Y, di Tomaso E, et al. Direct evidence that the VEGF-specific antibody bevacizumab has antivascular effects in human rectal cancer. *Nat Med* 2004;10:145–147.
94. Jain RK. Normalizing tumor vasculature with anti-angiogenic therapy: a new paradigm for combination therapy. *Nat Med* 2001;7:987–989.
95. Skliarenko JV, Lunt SJ, Gordon ML, et al. Effects of the vascular disrupting agent ZD6126 on interstitial fluid pressure and cell survival in tumors. *Cancer Res* 2006;66:2074–2080.
96. Raut CP, Morgan JA, Quek RH, et al. Measurement of interstitial fluid pressure (IFP) and circulating biomarkers in soft tissue sarcoma (STS): an exploratory phase II clinical and correlative study of sorafenib (SOR) in patients with refractory STS (NCI Protocol 6948). *J Clin Oncol* 2010;28:abstract 10091 (ASCO Meeting Abstracts).
97. Tailor TD, Hanna G, Yarmolenko PS, et al. Effect of pazopanib on tumor microenvironment and liposome delivery. *Mol Cancer Ther* 2010;9:1798–1808.
98. Ma J, Waxman DJ. Modulation of the antitumor activity of metronomic cyclophosphamide by the angiogenesis inhibitor axitinib. *Mol Cancer Ther* 2008;7:79–89.
99. Desai I, Van Laarhoven H, Hambrock T, et al. Assessment of early vascular effects of the angiogenesis inhibitor sunitinib (SU) in renal cell carcinoma (RCC) by dynamic contrast enhanced MRI (DCE-MRI) and diffusion weight MRI (DWI) at 3 tesla (T). *J Clin Oncol* 2010;28:abstract 3051 (ASCO Meeting Abstracts).
100. Crabb SJ, Patsios D, Sauerbrei E, et al. Tumor cavitation: impact on objective response evaluation in trials of angiogenesis inhibitors in non-small-cell lung cancer. *J Clin Oncol* 2009;27:404–410.
101. Dahlberg SE, Sandler AB, Brahmer JR, et al. Clinical course of advanced non-small-cell lung cancer patients experiencing hypertension during treatment with bevacizumab in combination with carboplatin and paclitaxel on ECOG 4599. *J Clin Oncol* 2010;28:949–954.
102. Hanrahan EO, Ryan AJ, Mann H, et al. Baseline vascular endothelial growth factor concentration as a potential predictive marker of benefit from vandetanib in non-small cell lung cancer. *Clin Cancer Res* 2009;15:3600–3609.
103. Bass MB, Sherman SI, Schlumberger MJ, et al. Biomarkers as predictors of response to treatment with motesanib in patients with progressive advanced thyroid cancer. *J Clin Endocrinol Metab* 2010;95:5018–5027.
104. Zhang W, Dahlberg SE, Yang D, et al. Genetic variants in angiogenesis pathway associated with clinical outcome in NSCLC patients (pts) treated with bevacizumab in combination with carboplatin and paclitaxel: Subset pharmacogenetic analysis of ECOG 4599. *J Clin Oncol* 2009;27:abstract 8032 (ASCO Meeting Abstracts).

105. Wilson PM, Yang D, Shi MM, et al. Use of intratumoral mRNA expression of genes involved in angiogenesis and HIF1 pathway to predict outcome to VEGFR tyrosine kinase inhibitor (TKI) in patients enrolled in CONFIRM1 and CONFIRM2. *J Clin Oncol* 2008;26: abstract 4002 (ASCO Meeting Abstracts).
106. Liu G, Rugo HS, Wilding G, et al. Dynamic contrast-enhanced magnetic resonance imaging as a pharmacodynamic measure of response after acute dosing of AG-013736, an oral angiogenesis inhibitor, in patients with advanced solid tumors: results from a phase I study. *J Clin Oncol* 2005;23:5464–5473.
107. Brahmer JR, Dahlberg SE, Gray RJ, et al. Sex differences in outcome with bevacizumab therapy: analysis of patients with advanced-stage non-small cell lung cancer treated with or without bevacizumab in combination with paclitaxel and carboplatin in the Eastern Cooperative Oncology Group Trial 4599. *J Thorac Oncol* 2011;6:103–108.
108. Sandler A, Yi J, Dahlberg S, et al. Treatment outcomes by tumor histology in Eastern Cooperative Group Study E4599 of bevacizumab with paclitaxel/carboplatin for advanced non-small cell lung cancer. *J Thorac Oncol* 2010;5:1416–1423.
109. Scartozzi M, Galizia E, Chiorrini S, et al. Arterial hypertension correlates with clinical outcome in colorectal cancer patients treated with first-line bevacizumab. *Ann Oncol* 2009;20:227–230.
110. Rixe O, Billefont B, Izzedine H. Hypertension as a predictive factor of sunitinib activity. *Ann Oncol* 2007;18:1117.
111. Friberg G, Kasza K, Vokes EE, et al. Early hypertension (HTN) as a potential pharmacodynamic (PD) marker for survival in pancreatic cancer (PC) patients (pts) treated with bevacizumab (B) and gemcitabine (G). *J Clin Oncol* 2005;23:abstract 3020 (ASCO Meeting Abstracts).
112. Hurwitz H, Douglas PS, Middleton JP, et al. Analysis of early hypertension (HTN) and clinical outcome with bevacizumab (BV). *J Clin Oncol* 2010;28:abstract 3039 (ASCO Meeting Abstracts).
113. Condeelis J, Pollard JW. Macrophages: obligate partners for tumor cell migration, invasion, and metastasis. *Cell* 2006;124:263–266.
114. Iyer AK, Khaled G, Fang J, et al. Exploiting the enhanced permeability and retention effect for tumor targeting. *Drug Discov Today* 2006;11: 812–818.